

Effects of Adolescent Caffeine Consumption on Cocaine Sensitivity

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Caffeine is the most commonly used psychoactive substance, and consumption by adolescents has risen markedly in recent years. We identified the effects of adolescent caffeine consumption on cocaine sensitivity and determined neurobiological changes within the nucleus accumbens (NAc) that may underlie caffeine-induced hypersensitivity to cocaine. Male Sprague-Dawley rats consumed caffeine (0.3 g/l) or water for 28 days during adolescence (postnatal day 28–55; P28–P55) or adulthood (P67–P94). Testing occurred in the absence of caffeine during adulthood (P62–82 or P101–121). Cocaine-induced and quinpirole (D₂ receptor agonist)-induced locomotion was enhanced in rats that consumed caffeine during adolescence. Adolescent consumption of caffeine also enhanced the development of a conditioned place preference at a sub-threshold dose of cocaine (7.5 mg/kg, i.p.). These behavioral changes were not observed in adults consuming caffeine for an equivalent period of time. Sucrose preferences were not altered in rats that consumed caffeine during adolescence, suggesting there are no differences in natural reward. **Caffeine consumption during adolescence reduced basal dopamine levels and augmented dopamine release in the NAc in response to cocaine (5 mg/kg, i.p.).** Caffeine consumption during adolescence also increased the expression of the dopamine D₂ receptor, dopamine transporter, and adenosine A₁ receptor and decreased adenosine A_{2A} receptor expression in the NAc. Consumption of caffeine during adulthood increased adenosine A₁ receptor expression in the NAc, but no other protein expression changes were observed. Together these findings suggest that caffeine consumption during adolescence produced changes in the NAc that are evident in adulthood and may contribute to increases in cocaine-mediated behaviors.

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INTRODUCTION

Caffeine is the most commonly used psychostimulant in the world (Rath, 2012). Caffeine is found naturally in coffee, tea and chocolate and is increasingly added as a supplement to other beverages including soda and energy drinks. Consumption of caffeine among adolescents has risen markedly in recent years in that daily caffeine consumption in 9- to 17-year olds has more than doubled since 1980 (Frary *et al*, 2005). Caffeine intake is positively correlated with substance-use disorders (Kendler *et al*, 2006), illicit drug use and other risky behaviors in young adults (Miller, 2008). Although moderate caffeine consumption in adults is considered relatively safe, there has been little research examining the long-term consequences of caffeine on the behavioral and neurobiological systems associated with substance use (Temple, 2009).

The psychostimulant actions of caffeine primarily result from nonselective blockade of both adenosine A₁ and A_{2A}

receptors throughout the brain (Fredholm *et al*, 1999). Adenosine A₁ and A_{2A} receptors are localized in the brain with very dense expression in striatal areas such as the nucleus accumbens (NAc) and caudate-putamen. Blockade of adenosine receptors within the striatal regions amplifies dopamine neurotransmission (Fuxe *et al*, 2007). **Caffeine increases extracellular dopamine in the striatum and prefrontal cortex (Borycz *et al*, 2007; De Luca *et al*, 2007; Solinas *et al*, 2002) and dopamine receptor antagonism blocks caffeine-induced locomotor activity (Garrett and Holtzman, 1994; Nehlig *et al*, 1994). Thus, caffeine's actions in the brain are likely due to both adenosine receptor antagonism as well as amplification of dopamine neurotransmission.**

The adolescent period is characterized by the maturation of brain systems including higher-order processing areas and a series of changes within the mesocorticolimbic dopamine system (Gladwin *et al*, 2011). **It is known that adolescents respond differently to caffeine compared with adults.** For example, adolescent rats are more sensitive to caffeine given they exhibit higher locomotor activation to caffeine compared with adults (Marin *et al*, 2011). Chronic caffeine consumption produces tolerance to caffeine to a greater extent in adolescents compared with adults, suggesting that adolescent animals may exhibit greater brain changes associated with chronic caffeine consumption

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(Rhoads *et al*, 2011). Chronic caffeine exposure in adolescents also produces behavioral cross-sensitization with methylphenidate when tested in adulthood in the absence of caffeine (Boeck *et al*, 2009). Given the differences in sensitivity and susceptibility adolescents display to the behavioral changes produced by chronic caffeine exposure, we hypothesized that caffeine consumption during adolescence may enhance the behavioral sensitivity to cocaine by producing changes in protein expression and dopamine release within the NAC.

MATERIALS AND METHODS

Animals

Male Sprague-Dawley rats (Charles River) were received on either postnatal day 21 (P21) or P60 and double housed with food and water *ad libitum*. All experimental procedures were conducted during the light period of a 12-h light/dark cycle and were completed in accordance with the guidelines established by the Institutional Animal Care and Use Committee at the University of Colorado Boulder.

Drugs

The nonselective adenosine receptor antagonist, caffeine, was purchased from Fisher Scientific (Waltham, MA). The dopamine D₂ receptor agonist, quinpirole ((-)-quinpirole hydrochloride), and cocaine hydrochloride were obtained from Sigma-Aldrich (St Louis, MO). All drugs, except caffeine, were dissolved in sterile-filtered physiological saline. Caffeine was dissolved in tap water.

Caffeine Consumption Procedure

Seven days after arrival, caffeine-consuming rats were given access to a single bottle containing caffeine in water (0.3 g/l) for 28 days (adolescent: P28–P55 or adult: P67–95; Figure 1). Age-matched control groups continued to receive water throughout the procedure. Caffeine and water consumption were monitored throughout the procedure. Following 28 days of caffeine exposure, the caffeine solution was replaced with water for the remainder of the experiment and behavioral testing was initiated at least 7 days after the last caffeine exposure. Thus, all behavioral testing and tissue collection were performed in the absence of caffeine between P62 and P82 or P101 and 121, periods corresponding to adulthood (Spear, 2000). Behavioral measures, tissue collection, and microdialysis studies were performed in separate cohorts of animals.

Locomotor Activity

Locomotor activity was conducted according to previously published procedures (Merritt and Bachtell, 2013). Briefly, animals underwent habituation to the locomotor chambers for 2 h on P62–65 (adolescent studies) or P101–104 (adult studies). On the following day, they were tested for cocaine- or quinpirole-induced locomotion in a single 4 h within-session escalating dose paradigm, where increasing doses of cocaine (vehicle, 2.5, 7.5 and 15 mg/kg, *i.p.*) or the dopamine D₂ receptor agonist, quinpirole, (vehicle, 0.1, 0.3

and 1.0 mg/kg, *i.p.*) were administered hourly. Locomotor activity was measured as the number of beam breaks during each hour of the testing period.

Place Conditioning

Place conditioning began 7 days following caffeine consumption (P62 for adolescent studies and P101 for adult studies) as described in the Supplementary Methods. Briefly, a three-phase procedure was conducted as follows: day 1—20 min pre-conditioning session, days 2–4—six 30 min conditioning sessions (0300 hours saline; 1500 hours cocaine) and day 5—20 min post-conditioning session. During the pre- and post-conditioning session, time spent in each compartment was recorded and the animals' preference was determined by subtracting the time in the drug-paired compartment from the time in the saline-paired compartment. We used 7.5 and 15 mg/kg cocaine to condition a place preference. The 7.5 mg/kg cocaine was chosen because our previous studies have demonstrated that it does not reliably produce a place preference in all rats (Merritt and Bachtell, 2013), making this cocaine dose useful in identifying differences in the development of a place preference between water- and caffeine-consuming groups.

Sucrose Preference

A two-bottle choice paradigm was used to test sucrose preferences. Seven days after caffeine removal (P62), rats were habituated to drink water from two bottles for 3 days. The experimental procedures were conducted over the subsequent 4 days where consumption was measured between 1800 and 2200 hours, a period corresponding to the onset of the dark cycle. On each day, the bottles were replaced during this period as follows: day 1—water/water, day 2—water/0.5% sucrose, day 3—water/water, and day 4—water/0.05% sucrose. Total consumption and a preference ratio ((sucrose consumption/total consumption) × 100) were used for analysis.

Dopamine Microdialysis

On P62 (7 days following caffeine exposure), unilateral microdialysis cannula (CMA Microdialysis, Solna, Sweden) were implanted under halothane anesthesia (1–2.5%) into either the right or left NAC shell (relative to bregma: AP = +1.7, ML = ±0.8, DV = -6.0) in a counterbalanced manner. Testing began ~1 week after recovery from surgical procedures (~P69). The evening before microdialysis testing, animals were transferred to the testing room and placed into separate plexiglass bowls containing bedding and *ad libitum* food and water. Microdialysis probes were inserted through the guide cannula and artificial CSF (145 mM NaCl, 2.7 mM KCl, 1.2 mM CaCl₂, and 1.0 mM KCl) was perfused through the probes overnight using a BASi infusion pumps at a flow rate of 0.2 µl/min overnight. The flow rate was increased to 1.5 µl/min the next morning. Following 90 min of equilibration, three 20 min baseline samples were collected for the first hour of the experiment. Before the fourth sample tube was inserted, rats received cocaine (5 mg/kg, *i.p.*). This dose produces a submaximal increase in extracellular dopamine in the NAC

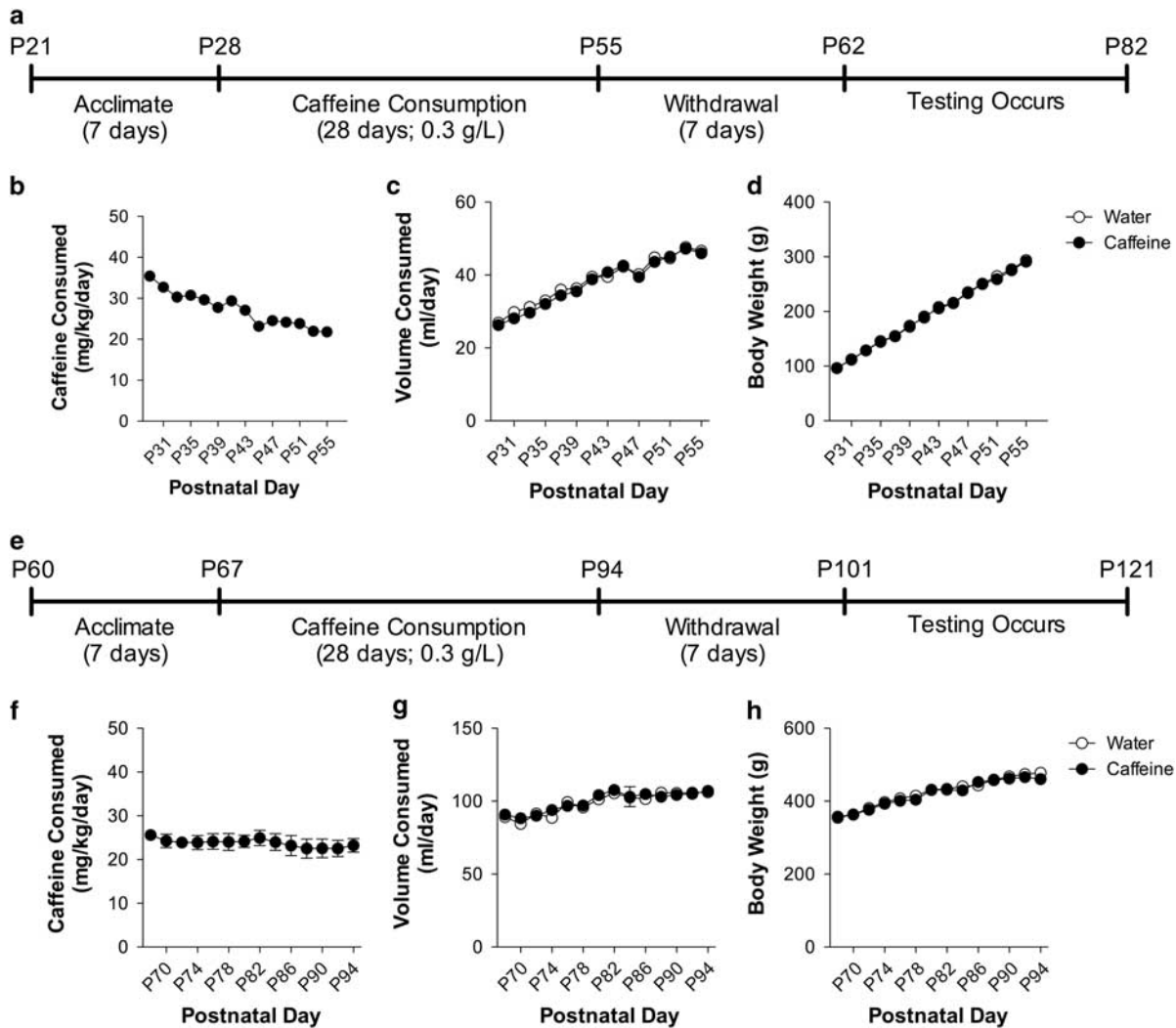


Figure 1 Caffeine consumption paradigm. (a) Time line of the model of adolescent caffeine consumption. Adolescent rats (P28–P55) consumed caffeine (0.3 g/l) and were tested in the absence of caffeine during adulthood (P62–P82). (b) Adolescent rats consumed an average of 27.34 ± 1.128 mg/kg/day, although the dose diminished over the adolescent period ($F_{14,225} = 130.2, p < 0.0001$). (c) There were no differences in the volume of fluid consumed by the caffeine-consuming rats ($n = 16$) compared with the water controls ($n = 16$). (d) Adolescent rats from both groups gained weight equivalently over the course of the consumption procedures. (e) Time line of the model of adult caffeine exposure. Adult rats (P67–P95) consumed caffeine (0.3 g/l) and were tested in the absence of caffeine (P101–P121). (f) The caffeine consumed remained stable over the procedure resulting in an average 23.78 ± 0.24 mg/kg of caffeine consumed per day. (g) There are no differences in the volume of fluid consumed by the caffeine-consuming rats ($n = 10$) compared with the water controls ($n = 10$). (h) There were also no differences in weight gain in the caffeine-consuming rats compared with the water controls.

and enables the detection of enhanced sensitivity for dopamine release. Samples were taken every 20 min for 3 h (total of nine samples) after injections, as previous studies indicate that cocaine-induced dopamine increases resolve within this time frame. Dopamine was quantified using an HPLC with electrochemical detection (ESA-Dionex, Sunnyvale, CA) in samples from animals whose cannulae were verified with cresyl violet staining post-mortem to be located within bounds of the desired region. One animal was removed from the study due to inaccurate placement.

Immunoblotting

Seven days following caffeine consumption (adolescent studies: P62 or adult studies: P101), rats were killed by rapid decapitation and bilateral 1 mm^3 tissue punches were taken

from chilled tissue slices containing the NAc and the caudate–putamen. Tissue punches were homogenized immediately and stored at -80°C until protein levels were quantified by a Lowry protein assay. Samples ($15 \mu\text{g}/\text{well}$) from each animal were separated by SDS-PAGE and electrophoretically transferred to PVDF membranes. Blots were incubated with affinity-purified primary antibodies (see Supplementary Methods). All blots were stripped and re-probed for the loading control protein, β -tubulin. Secondary antibodies were detected by enhanced chemiluminescence (ECL film) and densitized. Blots were run with equal numbers of water-exposed control and caffeine-exposed samples per gel and loaded in an alternating manner. The results were quantified using ImageJ and the optical density for the proteins was normalized to β -tubulin.

Data Analysis

The effects of caffeine consumption on the various behavioral and neurobiological outcomes were analyzed separately for adolescent and adult consumption studies. Body weight and consumption data (mg/kg/day and ml/day) were analyzed using a two-way mixed-design ANOVA with consumption group (between) and days (within) as factors. Locomotor data were analyzed using a two-way mixed-design ANOVA with consumption group (between) and cocaine or quinpirole dose (within) as factors. Place conditioning data were analyzed using a two-way between-subject ANOVA with consumption group and cocaine dose as factors. Dopamine measures in the microdialysis experiments were analyzed with either an unpaired *t*-test to test for consumption group differences in basal dopamine or a two-way mixed-design ANOVA for cocaine-induced dopamine release with consumption group (between) and time (within) as factors. Finally, effects of caffeine exposure on protein expression were analyzed separately using an unpaired *t*-test. In all cases, significant interactions and main effects were followed by planned comparisons using one-way ANOVA or Bonferroni's correction.

RESULTS

Caffeine Consumption During Adolescence

Throughout the caffeine-drinking procedure (Figure 1), caffeine consumption (mg/kg/day), the volume of fluid consumed per day (ml/day), and body weights (g) were recorded. Figure 1 displays data from one cohort of adolescents ($n = 32$) and adult animals ($n = 20$) used for cocaine-induced locomotor testing. Caffeine consumption averaged 27.34 ± 1.13 mg/kg/day (Figure 1b) and 23.78 ± 0.24 mg/kg/day (Figure 1f) for adolescent and adult animals, respectively. No differences between the water and caffeine groups were observed in either volume of fluid consumed (Figure 1c and g) or body weight (Figure 1d and h). In adolescent animals, there was a significant increase in total fluid consumption (days: $F_{13,429} = 255.3$, $p < 0.0001$) and body weight (days: $F_{13,377} = 3421$, $p < 0.0001$) during the procedure. Likewise, adult animals also showed a significant increase in total fluid consumption (days: $F_{13,112} = 14.11$, $p < 0.0001$) and body weight (days: $F_{13,252} = 491.8$, $p < 0.0001$) across the procedure. Analogous consumption data (data not shown) were collected for each cohort of adolescent and adult rats that were run through this procedure before examining all of the behavioral and neurobiological measures reported below.

Adolescent Caffeine Consumption Increases Cocaine Locomotor and Reward Sensitivity

To determine whether caffeine exposure during the adolescent period altered sensitivity to cocaine in adulthood, we examined locomotion to escalating doses of cocaine (0, 2.5, 7.5 and 15 mg/kg, i.p.). Caffeine consumption during adolescence increased locomotion to 15 mg/kg cocaine in adulthood (Figure 2a; dose \times group: $F_{3,84} = 3.45$, $p < 0.05$). To determine whether this effect was specific for chronic caffeine consumption during the adolescent period or

whether caffeine consumption influenced cocaine sensitivity regardless of the developmental period, we performed an analogous experiment in which adult animals consumed caffeine and were tested 7 days after removal of caffeine. Caffeine consumption in adult animals did not influence subsequent cocaine-induced locomotor sensitivity (Figure 2b).

In a separate cohort of animals, we examined the effect of adolescent caffeine exposure on the rewarding properties of cocaine using a place-conditioning paradigm. The development of a cocaine-induced conditioned place preference (CPP) for 7.5 mg/kg cocaine was more pronounced in the animals that consumed caffeine during adolescence compared with controls ($F_{1,44} = 9.62$, $p < 0.01$). Development of cocaine-induced CPP for 15 mg/kg cocaine was equivalent between groups. Adult caffeine-consuming animals showed no evidence of enhanced sensitivity for cocaine-induced CPP at the dose (7.5 mg/kg) tested (Figure 2d).

Adolescent Caffeine Consumption has no Effect on Sucrose Consumption and Preferences

We also tested sucrose consumption and preference in animals that consumed caffeine during adolescence to identify if these effects generalized to preferences for non-drug rewards. All animals displayed a preference for the sucrose-containing solution, and there were no group differences indicating that caffeine consumption during adolescence did not influence taste sensitivity or the ability to experience reward more generally (Supplementary Results).

Adolescent Caffeine Consumption Increases Dopamine D₂ Receptor Sensitivity

Evidence suggests that the expression and sensitivity of the dopamine D₂ receptor may correspond to enhanced psychostimulant use (Merritt and Bachtell, 2013; Volkow et al, 2006). We investigated whether adolescent caffeine consumption influenced the sensitivity to the dopamine D₂ receptor agonist, quinpirole, using an escalating dose-response procedure. Caffeine-consuming animals displayed significantly higher locomotor activation at 1 mg/kg quinpirole (Figure 2e; dose \times group: $F_{3,81} = 4.27$, $p < 0.01$). Adult caffeine-consuming animals did not show alterations in dopamine D₂ receptor sensitivity (Figure 2f).

Adolescent Caffeine Consumption Alters Extracellular Dopamine Levels in the NAC

Given the differences in cocaine-induced locomotion, cocaine CPP and dopamine D₂ receptor sensitivity observed in animals exposed to caffeine during adolescence, we next examined the effects of adolescent caffeine consumption on both basal dopamine levels and cocaine-induced dopamine release (Figure 3). Analysis of basal dopamine levels measured 60 min before cocaine administration revealed a significant reduction in animals that consumed caffeine during adolescence compared with controls (Figure 3a; $t_{10} = 2.49$, $p < 0.05$). Cocaine-induced dopamine release was increased in all animals following the 5 mg/kg cocaine (time: $F_{12,61} = 15.04$, $p < 0.0001$). This increase was enhanced in animals that consumed caffeine during adolescence (treatment \times time: $F_{12,61} = 2.70$, $p < 0.01$) at 40 ($t_{61} = 4.68$, $p < 0.001$) and 60 min

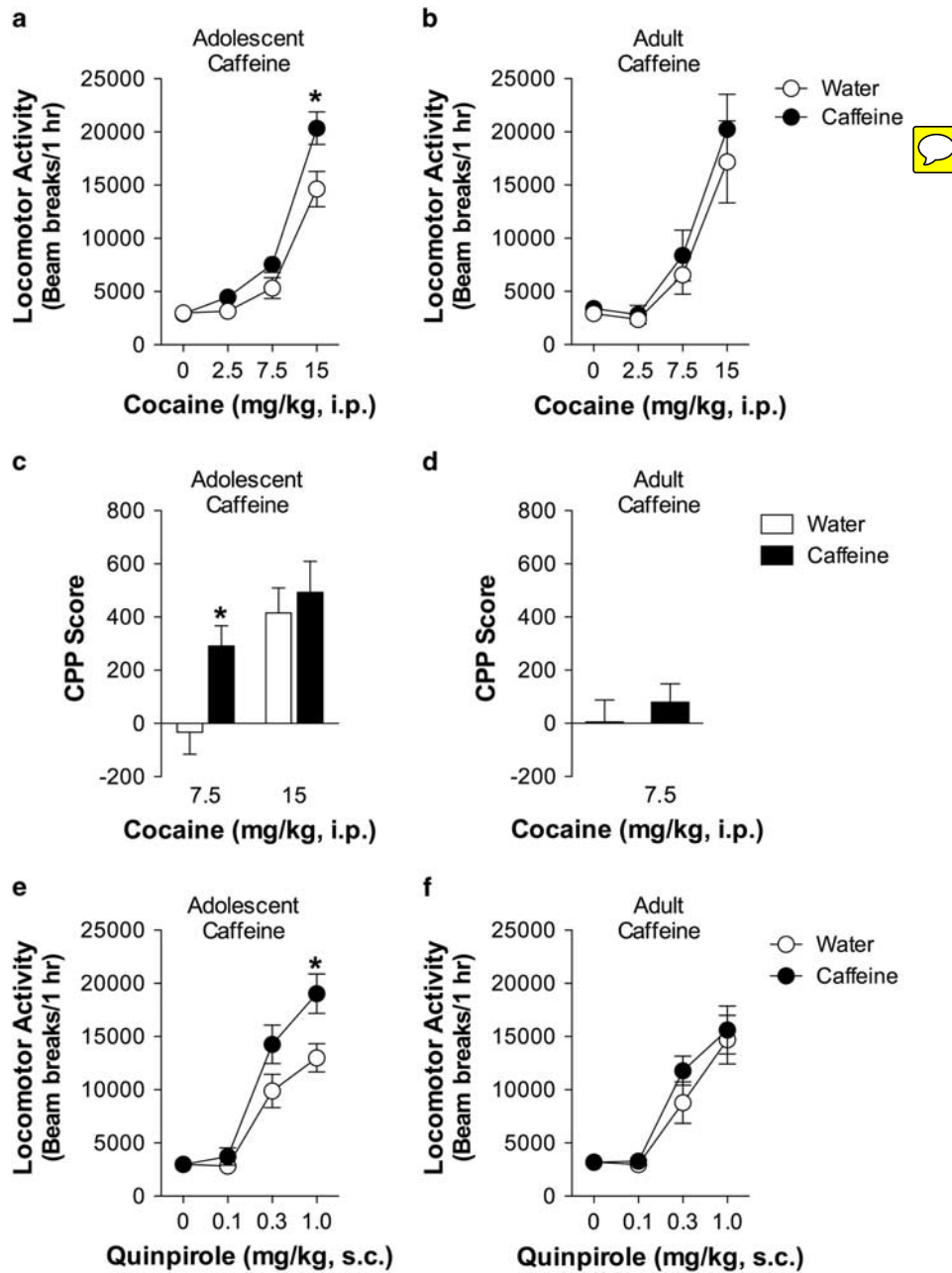


Figure 2 Caffeine consumption during adolescence enhances behavioral sensitivity to cocaine and quinpirole. (a) Rats were tested on an escalating dose regimen of cocaine (0, 2.5, 7.5 and 15 mg/kg, i.p.). Rats that consumed caffeine during adolescence ($n = 16$) display significantly higher locomotor activity to cocaine (15 mg/kg) compared with water controls ($n = 14$) following 7 days caffeine withdrawal in adulthood. * Significant from water controls ($t_{84} = 4.22$, $p < 0.001$). (b) Adult rats that consumed caffeine ($n = 8$) were not different in their cocaine sensitivity compared with water controls ($n = 8$). (c) Rats that consumed caffeine throughout adolescence ($n = 22$) develop a place preference at a sub-threshold dose of cocaine (7.5 mg/kg) compared with water controls ($n = 25$) following 7 days caffeine withdrawal in adulthood. * Significant from water controls ($p < 0.05$ Bonferroni's correction). Both groups (water: $n = 7$, caffeine: $n = 6$) developed CPP to 15 mg/kg cocaine, # $p < 0.05$ Bonferroni's correction. (d) Neither adult rats that consumed caffeine ($n = 17$) nor water controls ($n = 19$) developed a place preference to 7.5 mg/kg cocaine. (e) Rats were tested on an escalating dose regimen of quinpirole (0, 0.1, 0.3 and 1.0 mg/kg, s.c.). Rats that consumed caffeine during adolescence ($n = 15$) display significantly higher locomotor activity to 1.0 mg/kg quinpirole compared with water controls ($n = 15$) following 7 days caffeine withdrawal in adulthood. * Significant from water controls ($p < 0.05$ Bonferroni's correction). (f) Adult rats that consumed caffeine ($n = 8$) were not different in their quinpirole sensitivity compared with water controls ($n = 8$).

($t_{61} = 4.24$, $p < 0.01$) following cocaine administration (Figure 3b). Analysis of the raw dopamine dialysates before and after cocaine administration also revealed a significant difference between the groups ($F_{1,61} = 24.54$, $p < 0.001$). Only those rats that confirmed dialysis probe placements in the NAc shell were included in the analysis (Figure 3c).

Adolescent Caffeine Consumption Alters Protein Expression in the NAc

We next investigated the effects of chronic caffeine consumption during adolescence on several markers of adenosine and dopamine signaling in the NAc that could

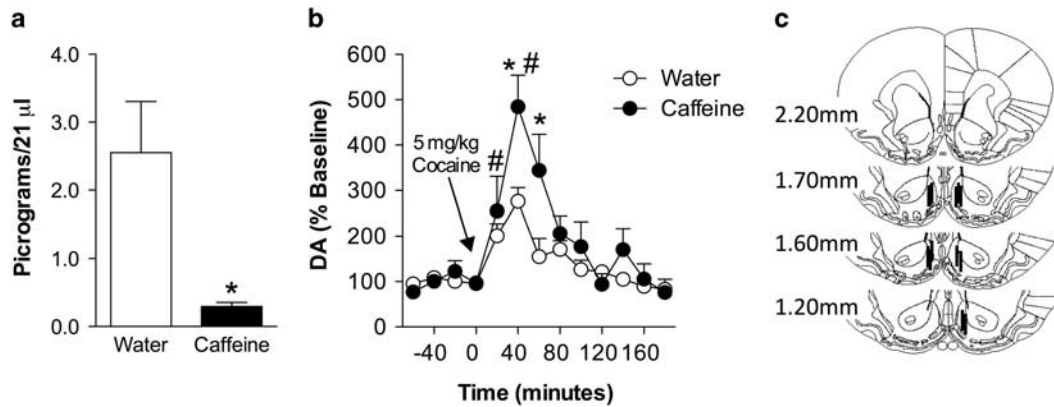


Figure 3 Caffeine consumption during adolescence lowers basal dopamine levels and increases cocaine-induced dopamine release in the nucleus accumbens (NAc). (a) Caffeine consumption during adolescence ($n=5$) reduced basal extracellular dopamine following 7 days caffeine withdrawal in adulthood (approximately P72) compared with water-consuming controls ($n=7$). (b) Both caffeine-consuming and water control animals exhibited a significant increase in dopamine following 5 mg/kg cocaine ($^{\#}p<0.05$ Bonferroni's correction compared with baseline) at 20 and 40 min time points. Rats consuming caffeine during adolescence displayed significantly enhanced cocaine-induced dopamine release. (c) Histological plates illustrating the accurate probe placements in the shell region of the NAc. * Significant from water controls ($p<0.05$ Bonferroni's correction).

contribute to the behavioral and neurochemical effects of caffeine. Protein expression was measured in tissue punches taken from the NAc and CPU 7 days following removal of caffeine (adolescent studies: P62, adult studies: P101). We observed a significant increase in adenosine A_1 receptor expression (Figure 4a; $t_{14}=2.37$, $p<0.05$) and a decrease in adenosine A_{2A} receptor expression (Figure 4a; $t_{14}=4.46$, $p<0.001$) in the NAc of animals that consumed caffeine during adolescence. Dopamine D_1 receptor expression was unaltered in the NAc between caffeine-exposed animals and water controls (Figure 4b), however, dopamine D_2 receptors were significantly enhanced in animals that consumed caffeine during adolescence (Figure 4b; $t_{14}=5.82$, $p<0.0001$). We also assessed the expression of presynaptic markers of dopamine neurotransmission, dopamine transporter (DAT), and tyrosine hydroxylase (TH) in the NAc. Caffeine consumption during adolescence increased DAT expression (Figure 4c; $t_{14}=3.37$, $p<0.01$) and did not change TH expression (data not shown). Finally, we measured total DARPP-32 expression as a marker for NAc medium spiny neurons. DARPP-32 expression was significantly enhanced by adolescent caffeine consumption ($t_{14}=3.00$, $p<0.01$). To identify whether these protein alterations were specific to the NAc, protein expression in the CPU was also examined. We found no significant differences in the protein expression of animals that consumed caffeine during adolescence in this brain region when compared with water controls (Supplementary Table 1). Similar to adolescent caffeine exposure, caffeine exposure during adulthood produced a significant increase in adenosine A_1 receptor expression in the NAc ($t_{14}=3.97$, $p<0.05$), but all other proteins were unchanged (Table 1).

DISCUSSION

Caffeine increases the rewarding effects of cocaine through its ability to increase dopamine neurotransmission (Bedingfield *et al*, 1998; Horger *et al*, 1991; Misra *et al*, 1986; Solinas *et al*, 2002). We examined the effects of

caffeine consumption during adolescence on cocaine-mediated behaviors and NAc neurobiology in adulthood following withdrawal from caffeine. Our findings reveal that adolescent caffeine exposure heightens sensitivity to cocaine-induced locomotion and facilitates development of cocaine-induced CPP. Sucrose preference and consumption were not altered by caffeine exposure, suggesting that these changes are specific to drug-related rewards. Adolescent consumption of caffeine also produced alterations in basal and cocaine-induced dopamine release in the NAc. We also observed both enhanced behavioral sensitivity to the dopamine D_2 receptor agonist, quinpirole, and enhanced expression of the D_2 receptor in the NAc. Finally, we identified increased adenosine A_1 receptor expression and decreased adenosine A_{2A} receptor expression in the NAc. Interestingly, adult animals that consumed caffeine for the same length of time did not show the same profile of behavioral or neurobiological alterations, suggesting that the developmental period of adolescence is especially sensitive to caffeine's effects on dopamine signaling in the NAc.

The increase in locomotor sensitivity to cocaine and quinpirole as well as the enhanced development of cocaine CPP may be mediated by the increases in dopamine D_2 receptor expression observed in the NAc. Previous work has shown that dopamine D_2 receptors are necessary for the development of cocaine sensitization (Fontana *et al*, 1993), and that increased sensitivity of dopamine D_2 receptors results in the development of cocaine CPP at a sub-threshold dose of cocaine (Merritt and Bachtell, 2013). It is unclear whether the increase in dopamine D_2 receptors reflects changes in presynaptic D_2 receptors on dopamine terminals, the postsynaptic D_2 receptors on medium spiny neurons, or both. Quinpirole administration produces a biphasic locomotor response (Herrera-Marschitz *et al*, 1988; Vanattou-Saifoudine *et al*, 2011; Zetterstrom and Fillenz, 1990). Low doses of quinpirole inhibit locomotion by stimulating high-affinity D_2 autoreceptors on dopamine terminals, while higher quinpirole doses increase locomotor activity following the saturation of D_2 autoreceptors and subsequent stimulation of postsynaptic D_2 receptors. We

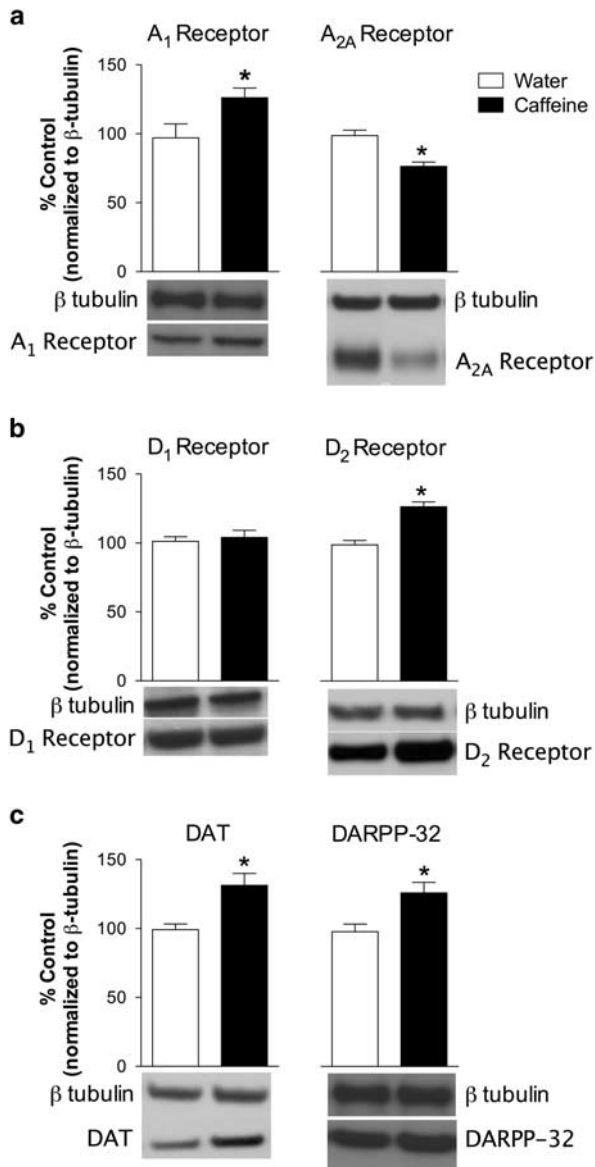


Figure 4 Caffeine consumption during adolescence produces alterations in markers of adenosine and dopamine signaling in the nucleus accumbens (NAc) during adulthood. (a) Adenosine A₁ receptor expression is increased while adenosine A_{2A} receptor expression is decreased in the NAc following adolescent consumption of caffeine. (b) Dopamine D₁ receptor expression is not altered by adolescent caffeine consumption, but dopamine D₂ receptor expression is significantly increased in the NAc following adolescent caffeine consumption. (c) Both the presynaptic marker of dopamine terminals, dopamine transporter (DAT), and postsynaptic marker of medium spiny neurons, DARPP-32, are significantly increased in the NAc following adolescent caffeine consumption compared with controls. *Significant from water control ($p < 0.05$ *t*-test), $n = 8$ /group.

observed enhanced quinpirole sensitivity at the high, postsynaptic doses, suggesting that the increases in dopamine D₂ receptors are postsynaptic. Previous work has suggested that increasing signaling in postsynaptic dopamine D₂ receptors enhances cocaine-related behaviors like reinstatement (Bachtell *et al*, 2005; Self *et al*, 1996), and that decreases, not increases, in dopamine D₂ autoreceptors lead to enhanced cocaine sensitivity (Bello *et al*, 2011).

Table 1 Effects of Adult Caffeine Consumption on Protein Expression in the Nucleus Accumbens

	Control (n = 8)	Caffeine (n = 8)
<i>Nucleus accumbens</i>		
Adenosine A ₁	100.0 ± 5.0	166.5 ± 14.3 ^a
Adenosine A ₂	100.0 ± 11.8	107.5 ± 7.4
Dopamine D ₁	100.0 ± 10.0	116.1 ± 9.8
Dopamine D ₂	100.0 ± 12.5	115.3 ± 8.7
Dopamine transporter	100.0 ± 14.9	113.9 ± 11.0
Tyrosine hydroxylase	100.0 ± 10.0	97.1 ± 10.4
DARPP-32	100.0 ± 6.6	109.0 ± 8.2

Values that are reported were normalized to b-tubulin and are expressed as % of water control.

^a $t_{1,4} = 3.97$, $p < 0.05$.

The enhanced activity of postsynaptic dopamine D₂ receptors coupled with the decreased expression of adenosine A_{2A} receptors in the NAc likely work in concert to amplify cocaine reward and sensitivity. Pharmacological experiments have shown that blockade of adenosine A_{2A} receptors in the NAc can increase locomotor sensitivity to cocaine (Filip *et al*, 2006), as well as cocaine seeking (O'Neill *et al*, 2014; Wu *et al*, 2010). Adenosine A_{2A} receptors exert tonic inhibitory control over D₂ receptor signaling within the striatum where A_{2A} receptor stimulation decreases DA binding at D₂ receptors (Ferre *et al*, 1991). Caffeine-induced enhancements in dopamine D₂ receptor expression coupled with diminished adenosine A_{2A} receptor expression may disengage adenosine's endogenous antagonism over D₂-mediated signaling. The increases in total DARPP-32 expression in the NAc also suggest increased dopamine signaling in the NAc that is associated with heightened sensitivity and reward to cocaine (Fienberg *et al*, 1998; Zachariou *et al*, 2002). This finding corroborates with a recent study where adolescent caffeine exposure increased both methylphenidate-induced locomotion and total DARPP-32 expression (Boeck *et al*, 2009). Together, these findings suggest that chronic caffeine consumption during adolescence enhances dopamine receptor signaling in the NAc.

Enhanced dopamine receptor signaling was also accompanied with alterations in extracellular dopamine concentrations in the NAc. Basal extracellular dopamine was significantly lower in the animals exposed to caffeine during adolescence. This is likely associated with a number of other changes observed in the NAc. For example, increases in DAT may translate into lower basal dopamine release in caffeine-exposed rats (Bello *et al*, 2011; Brodnik *et al*, 2013; Donovan *et al*, 1999). Genetic overexpression of DAT on dopamine neurons in the striatum produces decreased basal dopamine levels, as well as behavioral observations similar to our findings, such as increased cocaine-induced locomotion and increased development of cocaine CPP (Donovan *et al*, 1999; Salahpour *et al*, 2008). Furthermore, adenosine A₁ receptors are expressed on ~30% of dopamine terminals and act to inhibit dopamine release (Ebstein and Daly, 1982; Jin *et al*, 1993; Zetterstrom and Fillenz, 1990). We observed

an increase in expression of adenosine A₁ receptors that may provide excessive inhibition on the dopamine terminals. Interestingly, adult caffeine consumption also increased adenosine A₁ receptor expression in the NAC despite the fact that adult caffeine consumption did not recapitulate many of the behavioral changes observed following adolescent consumption. Additional studies should clarify the role of generalized adenosine A₁ receptor upregulation following caffeine consumption.

Although we observed decreases in basal dopamine levels, we found that systemic cocaine administration resulted in augmented dopamine release relative to baseline in animals exposed to caffeine during adolescence. It is not clear what, if any, changes in protein expression contribute to the enhanced dopamine release. Interestingly, we did not observe any change in TH, the enzyme associated with dopamine production, suggesting that there is no compensatory enhancement in the production of dopamine. The cortical areas that provide excitatory drive on the mesolimbic system are not fully developed at the time of the caffeine exposure (Pfeifer *et al*, 2011). It is possible that caffeine consumption alters the connectivity between the prefrontal cortex and the ventral tegmental area and/or the striatal areas. These alterations could indirectly influence phasic activation of the mesolimbic dopamine system and alter dopamine release. It is clear that this phenomenon should be examined more thoroughly to identify alterations in the ventral tegmental area and/or the cortical control regions to identify caffeine-induced abnormalities that would render an overactive dopamine system on cocaine administration.

Our studies suggest that, with the exception of the adenosine A₁ receptor upregulation, the behavioral and neurobiological changes observed following adolescent caffeine consumption are not observed in adults that consumed caffeine. These findings are in opposition to other studies where acute treatment with caffeine in adult animals increases the development of cocaine CPP (Bedingfield *et al*, 1998), augments cocaine-induced locomotor activity (Horger *et al*, 1991; Misra *et al*, 1986), and results in amplified activation of the mesolimbic dopamine system (Horger *et al*, 1991). Likewise, chronic caffeine exposure in adult animals increased locomotor activity to cocaine (Gasior *et al*, 2000; Schenk *et al*, 1990) and facilitated acquisition of cocaine self-administration (Carroll and Lac, 1998). Many of these studies use higher doses of caffeine as well as shorter exposure lengths both of which could produce different effects. In addition, and perhaps most importantly, all of these studies examined the animals' sensitivity to cocaine concurrent with caffeine administration. This suggests that, although chronic caffeine may enhance the psychostimulant effects of cocaine during the caffeine exposure, these effects are not long lasting and may 'washout' following withdrawal from caffeine. In contrast, it appears that the effects of chronic caffeine exposure during adolescence are prevalent during withdrawal from caffeine.

Caffeine intake is positively correlated with substance-use disorders (Kendler *et al*, 2006) and has been shown to increase illicit drug use and other risky behaviors in young adults (Miller, 2008). Despite the increase in caffeine consumption among adolescents, very few studies have examined the behavioral or neurobiological effects of

adolescent caffeine consumption. We provide new enlightening data suggesting that enduring alterations in reward pathway signaling are an important consequence of chronic adolescent caffeine consumption. On the basis of these findings, it is clear that more extensive studies are needed to determine caffeine's effects on brain development and possibly the permanent effects of adolescent caffeine exposure.

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The authors declare no conflict of interest.

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